



IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicants of: Voorberg, et al.

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Examiner: Maher M. Haddad

Filed: December 29, 2000

Docket: 294-86 PCT/US/RCE

For: METHOD FOR DIAGNOSIS AND
TREATMENT OF HEMOPHILIA A
PATIENTS WITH AN INHIBITOR

Dated: August 23, 2005

Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

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Dated: 8/23/05

DECLARATION UNDER 37 C.F.R. 1.132 OF JOHANNES JACOBUS

VOORBERG

Sir:

I, Johannes Jacobus Voorberg, Ph.D., declare as follows:

1. I am a co-inventor of the above-identified application.
2. I received a Ph.D. from the University of Amsterdam in 1991. I am an expert in the field of inhibitory antibodies directed to factor VIII. Currently, I am a senior scientist at Sanquin Research. Attached as Exhibit 1 is my *curriculum vitae*.

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3. I am a co-author of the Davies et al. reference (*Thromb. Haemostas.*, Supplement:2352, 1997) cited by the examiner in the Office Action of March 11, 2005. Davies et al. mentions that eight FVIII scFvs were selected by panning on immobilized rFVIII. In the Office Action, the examiner contends that the ability of the scFvs of Davies et al. to interfere with the activity of factor VIII inhibitors is an inherent property of the scFvs disclosed in Davies et al.
4. To demonstrate that the ability of a polypeptide to interfere with the activity of factor VIII inhibitors is not an inherent property of scFv polypeptides capable of specific binding to FVIII, the experiments described in exhibit 2 were performed by me or by persons directly under my supervision and control.
5. Briefly, a total of twenty different scFvs which bind to factor VIII were isolated. The twenty svFvs are listed in table 1 of exhibit 2. Twelve of the twenty scFvs were tested for their ability to neutralize (i.e., interfere with) factor VIII inhibitors. As shown in table 1, only five of the twelve tested scFvs were capable of neutralizing (i.e., interfere with) factor VIII inhibitors. See sample numbers 3, 16, 17, 18, and 19.
6. Therefore, not all svFv polypeptides that bind to factor VIII are capable of interference with factor VIII inhibitors. Accordingly, the ability of scFv to interfere with factor VIII inhibitors is not an inherent property of all scFvs which are capable of binding to factor VIII.

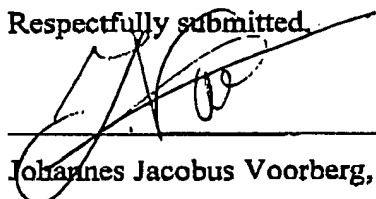
I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true, that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both under Section 1001 of Title 18 of

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the United States Code, and that such willful statements may jeopardize the validity of the application or any patent issued thereon.

Dated: 28/8/2005

Respectfully submitted,



Johannes Jacobus Voorberg, Ph.D.

Curriculum vitae Dr. J. Voorberg

Date of birth: 20-11-1961

University training: University of Wageningen, School of Molecular Sciences

Ph.D. training: Department of Molecular Biology under supervision of Prof. dr. H. Pannekoek en Dr. J.A. van Mourik (june 1987-june 1991)

Ph.D. thesis: "Biosynthesis and assembly of human von Willebrand factor" (Nov 1991) University of Amsterdam.

Current position: Senior Scientist at Department of Plasma Proteins, Sanquin Research, Amsterdam

Expertise: Biology of haemostatic proteins with emphasis on factor VIII and von Willebrand factor. Cell biology of von Willebrand factor in endothelial cells constitutes an important line of investigation. During the last five years research on molecular biological aspects of factor VIII inhibitor formation has been studied using phage display and transgenic animal models. These lines of research have resulted in over 40 publications and the following 4 Ph D theses:

"Consequences of haemophilia treatment" Karin Fijnvandraat (1998).

"Small GTP binding proteins involved in regulated secretion of von Willebrand factor from endothelial cells" Hubert P.J.C. de Leeuw (2000).

"Molecular analysis of factor VIII inhibitors using phage display" Edward N. van den Brink (2000).

"Inhibitor formation in mild haemophilia A" Wendy S. Bril (2003).

Foreign training: Massachusetts Institute of Technology, Boston
Department of Biochemistry (May-Dec, 1986)

University of Minnesota, Minneapolis
Department of Medicine, and Vascular Biology Center, University of Minnesota Medical School (Feb-July, 2004)

Research grants: Netherlands Organization of Scientific Research (NWO),
Netherlands Heart Foundation (NHS), Netherlands Thrombosis

Foundation (TSN), Landsteiner Foundation for Blood Transfusion
Research (LSBR) and Amsterdam-Leiden-Institute-for-
Immunology (ALIF).

List of publications

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2. Pannekoek, H. and **Voorberg, J.** (1989) Molecular cloning, expression and assembly of multimeric von Willebrand factor. *Baillieres Clin. Haematol.* 2, 879-896.
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4. Van de Ven, W.J., **Voorberg, J.**, Fontijn, R., Pannekoek, H., van den Ouweland, A.M., van Duijnhoven, H.L., Roebroek, A.J. and Siezen, R.J. (1990) Furin is a subtilisin-like proprotein processing enzyme in higher eukaryotes. *Mol. Biol. Rep.* 14, 265-270.
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Twenty scFvs that bind to factor VIII were isolated. The isolated scFvs' ability to inhibit factor VIII was determined as follows:

100 μ l of NP is diluted in 100 μ l 100 mM Tris-HCl pH=7.4; 0.4% HSA. Next, 200 μ l of different dilutions of scFv in 50 mM Tris-HCl pH=7.4; 0.2% HSA is added and incubated for 2 hour at 37°C. After incubation, residual factor VIII activity is determined is measured using a Sysmec C7000 (source of hemophilia A plasma: Dade Behring, OTXW17; Actin®FSL Activated PTT reagents; Dade Behring 52731334). Three different dilutions are measured per concentration of scFv. The ability of the scFvs to inhibit factor VIII is listed in table 1 below.

As shown in table 1, scFvs sample numbers 1, 5 and 7 inhibited factor VIII activity. The remaining scFvs samples (i.e., sample numbers 2-4, 6, 8-20) did not inhibit factor VIII activity.

The ability of scFv to interfere with the activity of factor VIII inhibitors was assayed as follows:

100 μ l of NP is diluted in 100 μ l 100 mM Tris-HCl pH=7.4; 0.4% HSA. Next, 150 μ l of different dilutions of scFv in 50 mM Tris-HCl pH=7.4; 0.2% HSA are added and for incubated for 1 hour at 37°C. After incubation, 50 μ l of diluted monoclonal antibody or diluted patient plasma is added till a final concentration of 2 BU/ml. The mixture is incubated for 1 hour at 37°C. Total volume is now 400 μ l. Sample is incubated for 1 hour and then residual factor VIII activity in the incubation mixture is measured using a Sysmec C7000 (source of hemophilia A plasma: Dade Behring, OTXW17; Actin®FSL Activated PTT reagents; Dade Behring 52731334). Three different dilutions are measured per concentration of scFv.

As shown in table 1, scFvs sample numbers 3, 16, 17, 18, and 19 are capable of interfering with factor VIII inhibitors, but do not inhibit factor VIII activity.

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Therefore, the ability of scFvs to interfere with factor VIII inhibitors is not an inherent property of scFvs to bind to factor VIII.

Table 1: Properties of factor VIII scFvs.

Sample No.	scFv Name	epitope	VH segment	Inhibition of Factor VIII	Interference with Factor VIII Inhibitors
1	VK34	A2	DP10	positive	negative
2	VK41	A2	DP47	negative	not tested
3	WP102	A2	DP47	negative	positive
4	WP137	A2	DP58	negative	not tested
5	B35	A3-C1	DP49	positive	negative
6	B04	A3-C1	DP77	negative	not tested
7	B38	A3-C1	DP15	positive	negative
8	KM37	A3-C1	DP14	negative	negative
9	KM38	A3-C1	DP49	negative	not tested
10	KM36	A3-C1	DP77	negative	not tested
11	WR1	C2	DP5	negative	negative
12	WR16	C2	DP5	negative	negative
13	WR17	C2	DP5	negative	negative
14	VK29	C2	DP25	negative	not tested
15	VK33	C2	DP79	negative	not tested
16	EL5	C2	DP14	negative	positive
17	EL14	C2	DP10	negative	positive
18	EL9	C2	DP88	negative	positive
19	IT2	C2	DP14	negative	positive
20	WR2	C2	DP88	negative	not tested

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